

Soluble non–starch polysaccharides affect net utilisation of energy by chickens

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Summary

Feed constituents such as the non–starch polysaccharides (NSPs) not only affect nutrient digestion in general, but also influence gut dynamics through changing the types and numbers of the microflora and the secretory response of the gut. The use of nutrients such as starch through microbial fermentation is not energy efficient; increased endogenous secretion of water, protein, lipids and minerals can affect post–digestive and, perhaps, post–absorptive processes because the gut secretes some 20 hormones or regulatory peptides. This paper discusses the importance of evaluating feed constituents from the standpoint of ‘efficiency’ and presents some experimental data on the effect of soluble NSPs on the net energy value and losses of energy as heat and as volatile fatty acids in the excreta of broiler chickens.

Introduction

The measures used to assess the nutritive value of poultry diets to date are metabolisable energy and amino acid contents, which also are the bases for practical diet formulations. The use of a net energy system for energy and digestible amino acid values for diet formulations has been strongly advocated in recent years (Farrell 1996; Ravindran *et al.* 1998; Ravindran and Bryden 1999). The rationale is that the efficiency of energy utilisation and the digestibility of amino acids are affected by diet constituents and that it is logical to consider the amounts of nutrients ‘available’ to the animal when evaluating feed. However, gut dynamics in relation to absorption of nutrients and their regulation and energetic costs have been given little attention in nutritional research. Considering the fact that 20% of whole animal energy expenditures can be accounted for by the gut (McBride and Kelly 1990; Cant *et al.* 1996), it is essential to investigate the way feed constituents behave in the gut and their effect on energetic efficiency of feed utilisation.

One constituent that has negative effect on the nutritive value of grains in poultry is the content of soluble non–starch polysaccharides (NSPs) (Burnett 1966; Antoniu *et al.* 1981; Annison 1991). This paper discusses how soluble NSPs affect energy utilisation beyond digestive processes in poultry.

Effect of NSPs on post–digestive processes

The effect of soluble NSPs on the apparent digestibility of nutrients in poultry has been studied extensively during the past ten years (Bedford *et al.* 1991; Choct and Annison 1992a). The negative correlation between NSPs in the diet and its nutritive value has been demonstrated in poultry (Choct and Annison 1990; Annison 1991), in pigs (King and Taverner 1975) and in dogs and cats (Earle *et al.* 1998). A general inhibition of the digestibility of nutrients occurs when diets with high levels of soluble NSPs are fed to chickens. Thus, Choct and Annison (1992a) demonstrated that addition of a soluble NSP isolate from wheat to a broiler diet depressed the ileal digestibilities of starch, protein and lipid by 14.6, 18.7 and 25.8%, respectively.

It is believed that viscous gut contents impede nutrient digestion by reducing the mixing of digestive enzymes with their substrates, but the effect of NSPs on the gut is not limited to their physical characteristics. They can induce changes to the microflora of the gut and modify endogenous secretion. These effects of NSPs on nutrient digestion are inter–related and depend on the solubility, molecular weights, and solution configurations of the polymers. Solubility of NSPs, in turn, depends on their chemical structures and association with other cell wall components.

The role of viscosity in the anti–nutritive effect of NSPs has been demonstrated by numerous workers (Bedford *et al.* 1991; Bedford and Classen 1992; Choct and Annison 1992b; Dusel *et al.* 1998; Steinfeldt *et al.* 1998). Choct and Annison (1992b) depolymerised an

isolate of wheat arabinoxylan (MW 758,000 Daltons) using a xylanase *in vitro* so that its viscosity was reduced four-fold. When these depolymerised NSPs were included in broiler diets, they did not exhibit strong anti-nutritive effects on nutrient digestion compared with intact NSPs included at the same level. NSPs can also bind nutrients and form complexes with digestive enzymes and some regulatory proteins in the gut. Angkanaporn *et al.* (1994) showed that addition of soluble arabinoxylans to a broiler diet markedly increased endogenous losses of amino acids, leading to a significant decrease in the apparent digestibility of protein in the ileum. The gut secretes some 20 hormones or regulatory peptides (Unväs-Moberg 1992), some enhance nutrient absorption, and others depress it. Feed components that have effects on endogenous protein secretion can be supposed to have an effect on hormonal secretions. Furthermore, viscous NSPs can enhance bile acid secretion and subsequently result in significant loss of these acids in the faeces (Ide *et al.* 1989; Ikegami *et al.* 1990). In addition to the modification of the gut physiology, certain NSPs can also bind bile salts, lipids and cholesterol (Vahouny *et al.* 1980; Vahouny *et al.* 1981). The net effect may be an altered lipid metabolism in the intestine, resulting in increased hepatic synthesis of bile acids from cholesterol to re-establish the composite pool of these metabolites in the enterohepatic circulation. The continued 'drain' of bile acids and lipids by sequestration, and increased elimination as faecal acidic and neutral sterols, may ultimately influence the absorption of lipids and cholesterol in the intestine. These effects could lead to major changes in the digestive and absorptive dynamics of the gut, with consequent poor overall efficiency in nutrient assimilation by the animal.

NSPs on energy losses via heat increment and as volatile substances in excreta

The energy derived from microbial fermentation in the chicken is small in quantity and inefficient as a metabolic fuel (Bolton and Dewar 1965). NSPs can bring about significant changes to the ecology of the gut, e.g. proliferation of fermentative microbes in the small intestine which ferment digestible nutrients such as starch to VFA (Choct *et al.* 1996). The amount of these VFA absorbed from the gut is not known. It is thought that the effect of soluble NSPs on the net utilisation of energy of feedstuffs for poultry may be more pronounced than that on the apparent metabolisable energy (AME) value because: (1) the digestive system of the bird would have to work harder than usual to cope with a highly viscous gut environment, and (2) the proliferation of fermentative organisms in the small intestine of chickens is energy-inefficient and detrimental to the bird. In a recent study using a close-circuit calorimetric system, we (Tukei, Thomson and

Choct, unpublished data) examined the effect of NSPs on energy utilisation in broiler chickens. In the first experiment, the loss of energy as VFA in the excreta of birds fed maize and barley was compared because these ingredients represent the two extremes amongst cereals in terms of their soluble NSP levels. As shown in Table 1, there was a large difference between birds fed maize and barley in the energy loss as VFA in the excreta, apparently indicating that elevated levels of soluble NSPs affected the way nutrients were digested. To test this finding further, we determined the net energy (NE) value of an NSP-enriched diet with or without enzyme supplementation in a second experiment. As the level of NSPs increased in the diet, the energy losses as heat increased and a considerable amount of energy was lost as VFA in the excreta. Thus when the anti-nutritive properties of the NSPs were removed by use of an exogenous enzyme, the AME and NE were increased by 29.1% and 37.3%, and heat production and energy loss as VFA were decreased by 11% and 61%, respectively. The data are shown in Table 2.

The increases in AME and NE were not proportional, indicating that NSPs not only interfere with digestive processes, but also have strong negative effects on net utilisation of energy. Part of this discrepancy may be explained by the loss of energy as VFA in the excreta. The use of nutrients through microbial conversion of digestible carbohydrates, such as starch, to VFA is not efficient compared to a direct absorption of glucose released from enzymatic digestion (Müller *et al.* 1989; Furuse and Okumura 1989; Carré *et al.* 1995). The production of VFA is energetically costly due to their costs of production and the energy losses as VFA in the excreta.

It is apparent that nutritionists should start to pay more attention to post-digestive and post-absorptive processes when evaluating nutritive quality. These include gut microbial balance, changes in gut dynamics such as secretory regulations and re-partitioning of nutrients. Muramatsu and colleagues (Muramatsu *et al.* 1983; 1987; 1993) clearly demonstrated that gut microflora increase energy costs by modifying the rate of energy-consuming reactions such as protein turnover within the chicken body. One such example is the gut cell turnover. According to LeBond and Walker (1956), a 100 g rat gaining 5 g/d synthesises 1 g mucosal cells daily, which represents a 20% additional tissue synthesis not manifest as weight gain. Extrapolating this to a 2 kg bird gaining 60 g daily, the bird would synthesise 12 g of mucosal tissue to maintain the integrity of its small intestine. Increased microbial load can exacerbate this loss (Abrams *et al.* 1963; Leshner *et al.* 1964) since some of its fermentation products, e.g. putrescine, have been shown to significantly enhance small intestinal and colonic mucosal growth rates (Osborne and Seidel 1989; Seidel *et al.* 1985). The indirect evidence of such costs is the often significant improvement in bird performance resulting from the inclusion of antibiotics in high-NSP diets (MacAuliffe

Table 1 Apparent metabolisable energy (AME) and loss of energy as VFA in the excreta of broilers fed maize or barley.

	Maize	Barley
AME (MJ/kg dry matter)	16.5	13.4
Excreta VFA energy (kJ/bird/day)	11.4	31.6
Excreta VFA energy (kJ/kg DM intake/bird)	159	433

Table 2 AME, net energy and loss of energy as VFA in the excreta of broilers fed a NSP enriched wheat based diet with or without enzymes.

	Wheat + NSPs (– enzyme)	Wheat + NSPs (+ enzyme)
AME (MJ/kg DM)	10.5	14.8
Respiratory quotient	0.995	1.006
Heat production (MJ/kg liveweight/day)	0.91	0.81
Net energy (MJ/kg DM)	4.7	7.5
Excreta VFA energy (kJ/bird/day)	56	17
Excreta VFA energy (kJ/kg DM intake/bird)	371	145

and McGinnis 1971; Misir and Marquardt 1978a,b). Supplementation with 150 mg procaine penicillin per kg of a diet consisting of 82% rye and 13.4% casein, resulted in increases of 75% in weight gain, 37% in feed intake and 37% in feed efficiency. However, there were no significant improvements in the AME (11.43 vs. 11.06 MJ/kg DM) and the digestibilities of starch (95% vs. 96%) and protein (40% vs. 40%) (Choct 1991). A better bird health and a more efficient nutrient utilisation are the likely reasons for this improvement.

Manipulation of efficiency of nutrient utilisation for poultry

In the light of a situation where fewer, if any, antibiotics may be allowed in feeds during the next decade, the use of antibiotic growth promotants is not a long term option for improving the efficiency of nutrient utilisation in the livestock industry. As shown in Table 2, the inclusion of appropriate glycanases in diets containing high levels of soluble NSPs can reduce energy losses as heat and as VFAs in the excreta, thus leading to improved net energy value. The reason for this action is speculative. Perhaps reducing the viscosity of the digesta in the small intestine hastens digesta passage and nutrient digestion rate (through removal of the diffusional constraint of viscous gums), thereby providing less substrate for the fermentative organisms and less time for them to proliferate. This may in turn promote the normal and efficient digestion (enzymatically) of starch and protein in the small intestine. Use of other

exogenous agents to manipulate gut dynamics and post-digestive and post-absorptive processes should be examined in a systematic manner for future feed and livestock production.

References

- Abrams, G.D., Bauer, H. and Sprinz, H. (1963). Influence of the normal flora on mucosal cell morphology and cellular renewal. *Laboratory Investigations* **12**, 355–363.
- Angkanaporn, K., Choct, M., Bryden, W.L., Annison, E.F. and Annison, G. (1994). Effects of wheat pentosans on endogenous amino acid losses in chickens. *Journal of the Science of Food and Agriculture* **66**, 399–404.
- Annison, G. (1991). Relationship between the levels of soluble non-starch polysaccharides and the apparent metabolizable energy of wheats assayed in broiler chickens. *Journal of Agriculture and Food Chemistry* **39**, 1252–1256.
- Antoniou, T., Marquardt, R.R. and Cansfield, E. (1981). Isolation, Partial characterization, and antinutritional activity of a factor (pentosans) in rye grain. *Journal of Agricultural and Food Chemistry* **28**, 1240–1247.
- Bedford, M. and Classen, H. (1992). Reduction of intestinal viscosity through manipulation of dietary rye and pentosanase concentration is effected through changes in the carbohydrate composition of the intestinal aqueous phase and results in improved growth rate and food conversion efficiency of broiler chicks. *Journal of Nutrition* **122**, 560–569.

- Bedford, M.R., Classen, H.L. and Campbell, G.L. (1991). The effect of pelleting, salt, and pentosanase on the viscosity of intestinal contents and the performance of broilers fed rye. *Poultry Science* **70**, 1571–1577.
- Bolton, W. and Dewar, W.A. (1965). The digestibility of acetic, propionic and butyric acids by the fowl. *Poultry Science* **56**, 2105–2107.
- Burnett, G.S. (1966). Studies of viscosity as the probable factor involved in the improvement of certain barleys for chickens by enzyme supplementation. *British Poultry Science* **7**, 55–75.
- Cant, J.P., McBride, B.W. and Croom, W.J., Jr. (1996). The regulation of intestinal metabolism and its impact on whole animal energetics. *Journal of Animal Science* **56**, 2541–2553.
- Carré, B., Gomez, J. and Chagneau, A.M. (1995). Contribution of oligosaccharide and polysaccharide digestion, and excreta losses of lactic acid and short chain fatty acids, to dietary metabolisable energy values in broiler chickens and adult cockerels. *British Poultry Journal* **36**, 611–629.
- Choct, M. (1991). The anti-nutritive effect of wheat pentosans in poultry. PhD thesis. University of Sydney.
- Choct, M. and Annison, G. (1990). Anti-nutritive activity of wheat pentosans in broiler diets. *British Poultry Science* **31**, 811–821.
- Choct, M. and Annison, G. (1992a). The inhibition of nutrient digestion by wheat pentosans. *British Journal of Nutrition* **67**, 123–132.
- Choct, M. and Annison, G. (1992b). Anti-nutritive effect of wheat pentosans in broiler chickens: roles of viscosity and gut microflora. *British Poultry Science* **33**, 821–834.
- Choct, M., Hughes, R.J., Wang, J., Bedford, M.R., Morgan, A.J. and Annison, G. (1996). Increased small intestinal fermentation is partly responsible for the anti-nutritive activity of non-starch polysaccharides in chickens. *British Poultry Science* **37**, 609–21.
- Dusel, G., Kluge, H. and Jeroch, H. (1998). Xylanase supplementation of wheat-based rations for broilers: Influence of wheat characteristics. *Journal of Applied Poultry Science* **7**, 119–131.
- Earle, K.E., Opitz, B., Kienzle, E., Smith, P.M. and Maskell, I.E. (1998). Effect of fiber on digestibility of organic matter and energy in pet foods. www.waltham.com/vets/pubs/vp02_14.htm
- Farrell, D.J. (1996). Energy systems for poultry: Recent Developments. *Proceedings of World's Poultry Science Congress*, pp. 225–235. New Delhi, India.
- Furuse, M and Okumura, J. (1989). Effects of acetic acid levels on protein and energy utilisation in chicks. *Poultry Science* **68**, 795–798.
- Ide, T., Horii, M., Kawashim, K. and Yamamoto, T. (1989). Bile acid conjugation and hepatic taurine concentration in rats fed on pectin. *British Journal of Nutrition* **62**, 539–550.
- Ikegami, S., Tsuchihashi, F., Harada, H., Tsuchihashi, N., Nishide, E. and Innami, S. (1990). Effect of viscous indigestible polysaccharides on pancreatic–biliary secretion and digestive organs in rats. *Journal of Nutrition* **120**, 353–360.
- King, R.H. and Taverner, M.R. (1975). Prediction of the digestible energy in pig diets from analyses of fibre contents. *Animal Production* **21**, 275–284.
- LeBond, C.P. and Walker, B.E. (1956). Renewal of cell populations. *Physiology Reviews* **36**, 255–276.
- Leshner, S., Wallbury, Jr. H.E. and Sacher, G.A. (1964). Generation cycle in the duodenal crypt cells of germ-free and conventional mice. *Nature* **202**, 884–886.
- MacAuliffe, T. and McGinnis, J. (1971). Effect of antibiotic supplements to diets containing rye on chick growth. *Poultry Science* **50**, 1130–1134.
- McBride, B.W. and Kelly, J.M. (1990). Energy cost of absorption and metabolism in the ruminant gastrointestinal tract and liver: A review. *Journal of Animal Science* **68**, 2997–3010.
- Misir, R. and Marquardt, R.R. (1978a). Factors affecting rye (*Secale cereale* L.) utilization in growing chicks. I. The influence of rye level, ergot and penicillin supplementation. *Canadian Journal of Animal Science* **58**, 691–701.
- Misir, R. and Marquardt, R.R. (1978b). Factors affecting rye (*Secale cereale* L.) utilisation in growing chicks. II. The influence of protein type, protein level and penicillin. *Canadian Journal of Animal Science* **58**, 703–715.
- Müller, H.L., Kirchgessner, M. and Roth, F.X. (1989). Energy utilisation of intracaecally infused carbohydrates and casein in sows. In: *Energy Metabolism of Farm Animals*, pp123–126. (eds. Y. van der Honing, and W.H. Close). Wageningen, Pudoc, The Netherlands.
- Muramatsu, T., Coates, M.E., Hewitt, D., Salter, D.N. and Garlick, P.J. (1983). The influence of the gut microflora on protein synthesis in liver and jejunal mucosa in chicks. *British Journal of Nutrition* **49**, 453–462.
- Muramatsu, T., Takasu, O., Furuse, M., Tasaki, I. and Okumura, J. (1987). Influence of gut microflora on protein synthesis in tissues and in the whole body of chicks. *Biochemical Journal* **246**, 475–479.
- Muramatsu, T., Takemura, J. and Okumura, J. (1993). Acetic acid is not involved in enhanced intestinal protein synthesis by the presence of the gut microflora in chickens. *Comparative Biochemistry and Physiology* **105A**, 543–548.
- Osborne, D.L. and Seidel, E.R. (1989). Microflora derived polymines modulate obstruction induced colonic mucosal hypertrophy. *American Journal of Physiology* **256**, G1049–G1057
- Ravindran, V. and Bryden, W.L. (1999). Evaluation of broiler diets containing graded levels of cottonseed meal and formulated on the basis of total or digestible amino acids. *Proceedings of the Australian Poultry Science Symposium* **11**, 168.

- Ravindran, V., Hew, L.I. and Bryden, W.L. (1998). Broiler feed formulations with canola meal based on total or digestible amino acids. *Proceedings of the Australian Poultry Science Symposium* **10**, 209.
- Seidel E.R, Haddox, M.K and Johnson, L.R. (1985). Ileal mucosal growth during intraluminal infusion of ethylamine or putrescine. *American Journal of Physiology* **249**, G434–G438.
- Steenfeldt, S., Hammershoj, M., Mullertz, A. and Jensen, J.F. (1998). Enzyme supplementation of wheat–based diets for broilers. 2. Effect on apparent metabolisable energy content and nutrient digestibility. *Animal Feed Science and Technology* **75**, 45–64.
- Unväs–Moberg, K. (1992). The endocrine system of the gut during growth and reproduction, role of afferent and efferent mechanisms. *Proceedings of the Nutrition Society of Australia* **17**, 167–176.
- Vahouny, G.V., Roy, T., Gallo, L.L., Story, J.A., Kritchevsky, D. and Cassidy, M. (1980). Dietary fibers 3 effects of chronic intake on cholesterol absorption and metabolism in the rat. *The American Journal of Clinical Nutrition* **33**, 2182–2191.
- Vahouny, G.V., Tombes, R., Cassidy, M.M., Kritchevsky, D. and Gallo, L.L. (1981). Dietary fibres. VI: Binding of fatty acids and monolein from mixed micelles containing bile salts and lecithin. *Proceedings of the Society of Experimental Biology and Medicine* **166**, 12–16.

